

# Corynebacterium glutamicum

Multiporator/Eppendorf Eporator®

## Transformation Protocol

Protocol No. 4308 915.510 – 12/2001

<b>Microorganism</b>	<i>Corynebacterium glutamicum</i>
<b>Cell type</b>	Bacteria, gram positive
<b>Molecules injected</b>	Plasmid DNA
<b>Growth medium</b>	LBG (LB with 0.5% glucose) supplemented with 2.5% glycine and isonicotinic acid hydrazid (4 mg/ml)
<b>Washing solution</b>	15% glycerol (v/v)
<b>Electroporation solution</b>	15% glycerol (v/v)
<b>Outgrowth medium</b>	LBG medium (LB with 0.5% glucose)
<b>Cuvette</b>	2 mm gap width
<b>Reference</b>	Haynes J. and Britz M. • 1990 • Journal of General Microbiology 136 • 255-263

### Making electrocompetent cells:

1. Grow cells in LBG medium at 30 °C and shaking at 200 rpm to an O.D.<sub>600</sub> of 0.15 to 0.25.
2. Harvest by centrifugation.
3. Wash in one culture volume of 15% glycerol.
4. Resuspend cells in 0.002 culture volumes of 15% glycerol (number of cells: approx.  $2.5 \times 10^{10}$  cells/ml).

### Electroporation of cells:

1. Add 1-2 µl plasmid DNA (in water) to a minimum of 40 µl of electrocompetent cells. Homogenize by gently mixing with pipette several times. Transfer mixture into a prechilled cuvette.
2. Wipe moisture from the cuvette and insert the cuvette into the device.
3. Electroporation:

<b>Mode</b>	Prokaryotes "O"
<b>Voltage (V)</b>	2,500 V
<b>Time constant (τ)</b>	5 ms

4. Immediately transfer cell suspension into LBG medium at a 1:25 dilution, incubate 1 hour at 30 °C.
5. Plate cells on selective plates; incubate for 4 days.

### Expected results:

Transformation efficiency up to  $5 \times 10^5$  transformants/µg of DNA.

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