

# Listeria monocytogenes

Multiporator/Eppendorf Eporator®

## Transformation Protocol

Protocol No. 4308 915.520 – 12/2001

<b>Microorganism</b>	<i>Listeria monocytogenes</i> 23074
<b>Cell type</b>	Bacteria, gram positive
<b>Molecules injected</b>	Plasmid DNA
<b>Growth medium</b>	BHI, supplemented with 0.5 M sucrose
<b>Washing solution</b>	1 mM HEPES (pH 7.0), 0.5 M sucrose
<b>Electroporation solution</b>	1 mM HEPES (pH 7.0), 0.5 M sucrose
<b>Outgrowth medium</b>	BHI, supplemented with 0.5 M sucrose
<b>Cuvette</b>	1 mm gap width
<b>Reference</b>	Park, S. and Stewart, G. • 1990 • Gene 94 • 129-132

### Making electrocompetent cells:

1. Dilute an overnight culture of *L. monocytogenes* 23074 in BHI, into fresh media (1:100). Grow at 37 °C with shaking until reaching an O.D.<sub>600</sub> of 0.2.
2. Add Penicillin G (10 µg/ml) and continue incubation for a further 2 hours.
3. Harvest by centrifugation (8,000 x g, 10 min., at 4 °C) and wash three times in ice-cold washing solution, once with equal volume and twice with ½ volume.
4. Resuspend the cells in ice-cold electroporation solution (0.0025 vol.). Use cells within 30 minutes of their preparation.

### Electroporation of cells:

1. Add 1 µg plasmid DNA to 100 µl of electrocompetent cells. Homogenize by gently mixing with pipette several times. Transfer mixture into a prechilled cuvette.
2. Wipe moisture from the cuvette and insert the cuvette into the device.
3. Electroporation:

<b>Mode</b>	Prokaryotes "O"
<b>Voltage (V)</b>	1,000 V
<b>Time constant (τ)</b>	5 ms

4. Immediately add 1 ml of BHI medium and incubate at 37 °C for one hour.
5. Plate cells on selective BHI plates.

### Expected results:

Transformation efficiency up to 4 x 10<sup>6</sup> transformants/µg of DNA.

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