

# *Serratia liquefaciens*

Multiporator/Eppendorf Eporator ®

Transformation Protocol

Protocol No. 4308915.550

<b>Microorganism</b>	<i>Serratia liquefaciens</i>
<b>Cell type</b>	Bacteria, gram negative
<b>Molecules injected</b>	Plasmid DNA pDsRed
<b>Growth medium</b>	LB medium
<b>Washing solution</b>	10% glycerol
<b>Electroporation solution</b>	10% glycerol
<b>Outgrowth medium</b>	LB medium with antibiotics agar plates
<b>Cuvette</b>	1 mm gap width
<b>Reference</b>	<b>Lilian Pilares and Jose Ramos Vivas, IFIMAV, Santander, Spain.</b>

## Making electrocompetent cells:

1. Dilute an overnight culture of *S. liquefaciens* in LB, into fresh media (1:100). Grow at 37 °C with shaking until reaching an O.D.<sub>600</sub> of 0.5.
2. Chill cells on ice for 10 minutes and transfer to a pre-chilled centrifuge tubes to harvest cells. Wash three times with cold sterile 10% glycerol. The first time in 25 ml, the second time in 12,5 ml, the third and fourth times in 2 ml.
3. Resuspend in 10% glycerol at a concentration of approx. 10<sup>9</sup> cells/ml and store at -70 °C until needed.

## Electroporation of cells:

1. Add 1 µg plasmid DNA to 50 µl (~10<sup>9</sup>) of electro competent cells. Homogenize by gently mixing with pipette several times. Transfer mixture into a pre-chilled cuvette (-20°C).
2. Wipe moisture from the cuvette and insert the cuvette into the device.
3. Electroporation:

Voltage (V)	1,800 V
Time constant (t)	5ms
4. Immediately add 1 ml of pre-warmed LB medium and incubate at 37°C for 1 hour with strong shaking.
5. Plate on selective agar plates (ampicillin) and incubate al 37°C for 24 h.

## Expected results:

Transformation efficiency from 10<sup>5</sup> to 5×10<sup>7</sup> transformants/µg of DNA (strain-dependent differences).